ELSEVIER

Journal of Microbiological Methods 31 (1998) 209–210

Journal of Microbiological Methods

Author Index of Volume 31

Abalia, I., see Blanco, M. (1/2) 37
Alonso, M.P., see Blanco, M. (1/2) 37
Alugupalli, S., Sikka, M.K., Larsson, L. and White, D.C., Gas chromatography–mass spectrometry methods for the analysis of mycocerosic acids present in *Mycobacterium tuberculosis* (3) 143
Araujo, R., see Puig, A. (3) 205

Barbian, K.D., see Minnick, M.F. (1/2) 51
Bazin, M.J., see Pearce, D.A. (1/2) 67
Binnerup, S.J., Højberg, O. and Sørensen, J., Gram characteristics determined on single cells and at the microcolony level of bacteria immobilized on polycarbonate membrane filters (3) 185
Blanco, J., see Blanco, M. (1/2) 37
Blanco, J.E., see Blanco, M. (1/2) 37
Blanco, M., Blanco, J.E., Rodríguez, E., Abalia, I., Alonso, M.P. and Blanco, J., Detection of virulence genes in uropathogenic *Escherichia coli* by polymerase chain reaction (PCR): comparison with results obtained using phenotypic methods (1/2) 37
Bobish, J. and Leisner, S., Novel use of *polA* bacteria for inserting DNA fragments into *Agrobacterium* binary vectors (1/2) 89
Boccuzzi, V.M., Straube, W.L., Ravel, J., Colwell, R.R. and Hill, R.T., Preparation of DNA extracted from environmental water samples for PCR amplification (3) 193
Bracey, D., Holyoak, C.D., Nebe-von Caron, G. and Coote, P.J., Determination of the intracellular pH (pH_i) of growing cells of *Saccharomyces cerevisiae*: the effect of reduced-expression of the membrane H^+-ATPase (3) 113
Braux, A.-S., Minet, J., Tamanai-Shacoori, Z., Riou, G. and Cormier, M., Direct enumeration of injured *Escherichia coli* cells harvested onto membrane filters (1/2) 1
Brill, J.A. and Wiegel, J., Differentiation between spore-forming and asporogenic bacteria using a PCR and Southern hybridization based method (1/2) 29
Brun, F., see Zbinden, R. (1/2) 95

Clement, B.G., Kehl, L.E., DeBord, K.L. and Kitts, C.L., Terminal restriction fragment patterns (TRFPs), a rapid, PCR-based method for the comparison of complex bacterial communities (3) 135
Colwell, R.R., see Boccuzzi, V.M. (3) 193
Coote, P.J., see Bracey, D. (3) 113
Cormier, M., see Braux, A.-S. (1/2) 1
Cormiér, M.R., see Macnaughton, S.J. (1/2) 19
Correll, R.L., Harch, B.D., Kirkby, C.A., O'Brien, K. and Pankhurst, C.E., Statistical analysis of reduction in tensile strength of cotton strips as a measure of soil microbial activity (1/2) 9
Criado, M.T., see Echeverría, B. (3) 151
Cristani, C., see Vannacci, G. (3) 175

DeBord, K.L., see Clement, B.G. (3) 135
Dott, W., see Schmitz, R.P.H. (3) 159
Duffy, G. and Sheridan, J.J., Viability staining in a direct count rapid method for the determination of total viable counts on processed meats (3) 167

Echeverría, B., Gómez, J.A., Hernández, E., Criado, M.T. and Ferreirós, C., Optimization of membrane isolation and transferrin-binding proteins solubilization from *Neisseria meningitidis* cells (3) 151
Eisenträger, A., see Schmitz, R.P.H. (3) 159

Ferreirós, C., see Echeverría, B. (3) 151
Fremming, C., see Nexmann Jacobsen, C. (1/2) 75

Gómez, J.A., see Echeverría, B. (3) 151

Harch, B.D., see Correll, R.L. (1/2) 9
Henriksen, K., see Roslev, P. (3) 99
Hernández, E., see Echeverría, B. (3) 151
Hill, R.T., see Boccuzzi, V.M. (3) 193
Højberg, O., see Binnerup, S.J. (3) 185
Holyoak, C.D., see Bracey, D. (3) 113

Iversen, N., see Roslev, P. (3) 99

Jakobsen, M., see Nexmann Jacobsen, C. (1/2) 75
Jeleń, H., Latus-Ziętkiewicz, D., Wąsowicz, E. and Kamiński, E., Trichodiene as a volatile marker for trichothecenes biosynthesis (1/2) 45
Jenkins, T.L., see Macnaughton, S.J. (1/2) 19
Jofre, J., see Puig, A. (3) 205

Kamiński, E., see Jeleń, H. (1/2) 45
Kehl, L.E., see Clement, B.G. (3) 135
Kijima-Tanaka, M., Ogikubo, Y., Kojima, A. and Tamura, Y., Development of a two-site enzyme-linked immunosorbent assay for quantification of the flagellar antigen in blackleg vaccines (1/2) 83

Kirkby, C.A., see Correll, R.L. (1/2) 9
Kitts, C.L., see Clement, B.G. (3) 135
Kobayashi, R.K.T. and Vidotto, M.C., Digoxigenin-labelled probe for the detection of enteropathogenic *Escherichia coli* (3) 201
Kojima, A., see Kijima-Tanaka, M. (1/2) 83

Larsson, L., see Alugupalli, S. (3) 143
Latus-Ziętkiewicz, D., see Jeleń, H. (1/2) 45
Lee, H., see Tin Leung, K. (1/2) 59
Leisner, S., see Bobish, J. (1/2) 89
Lynch, J.M., see Pearce, D.A. (1/2) 67

Macnaughton, S.J., Jenkins, T.L., Wimpee, M.H., Cormiér, M.R. and White, D.C., Rapid extraction of lipid biomarkers from pure culture and environmental samples using pressurized accelerated hot solvent extraction (1/2) 19
Minet, J., see Braux, A.-S. (1/2) 1
Minnick, M.F. and Barbian, K.D., Identification of *Bartonella* using PCR; genus- and species-specific primer sets (1/2) 51
Müller, F., see Zbinden, R. (1/2) 95

Nakao, H. and Popovic, T., Development of a rapid ribotyping method for *Corynebacterium diphtheriae* by using PCR single-strand conformation polymorphism: comparison with standard ribotyping (3) 127
Nebe-von Caron, G., see Bracey, D. (3) 113
Nexmann Jacobsen, C., Fremming, C. and Jakobsen, M., Immunomagnetic separation of *Listeria monocytogenes* for flow cytometric determination of viable cells in liquid (1/2) 75

O'Brien, K., see Correll, R.L. (1/2) 9
Ogikubo, Y., see Kijima-Tanaka, M. (1/2) 83

Pankhurst, C.E., see Correll, R.L. (1/2) 9
Pearce, D.A., Bazin, M.J. and Lynch, J.M., A physical model system in which to investigate the interactions of microorganisms isolated from the rhizosphere (1/2) 67
Popovic, T., see Nakao, H. (3) 127
Puig, A., Jofre, J. and Araujo, R., Use of oxrase enzyme (Oxrase®) for the detection of bacteriophages of *Bacteroides fragilis* in aerobic incubating conditions (3) 205

Ravel, J., see Boccuzzi, V.M. (3) 193
Riou, G., see Braux, A.-S. (1/2) 1
Rodríguez, E., see Blanco, M. (1/2) 37
Roslev, P., Iversen, N. and Henriksen, K., Direct fingerprinting of metabolically active bacteria in environmental samples by substrate specific radiolabelling and lipid analysis (3) 99

Schmitz, R.P.H., Eisenträger, A. and Dott, W., Miniaturized kinetic growth inhibition assays with *Vibrio fischeri* and *Pseudomonas putida* (application, validation and comparison) (3) 159
Sheridan, J.J., see Duffy, G. (3) 167
Sikka, M.K., see Alugupalli, S. (3) 143
Sørensen, J., see Binnerup, S.J. (3) 185
Straube, W.L., see Boccuzzi, V.M. (3) 193

Tamanai-Shacoori, Z., see Braux, A.-S. (1/2) 1
Tamura, Y., see Kijima-Tanaka, M. (1/2) 83
Tin Leung, K., Watt, A., Lee, H. and Trevors, J.T., Quantitative detection of pentachlorophenol-degrading *Sphingomonas* sp. UG30 in soil by a most-probable-number/polymerase chain reaction protocol (1/2) 59
Trevors, J.T., see Tin Leung, K. (1/2) 59

Vannacci, G. and Cristani, C., Characterization of chlorate-resistant sectors from isolates of *Fusarium moniliforme* and *F. proliferatum* (3) 175
Vidotto, M.C., see Kobayashi, R.K.T. (3) 201
von Graevenitz, A., see Zbinden, R. (1/2) 95

Wąsowicz, E., see Jeleń, H. (1/2) 45
Watt, A., see Tin Leung, K. (1/2) 59
White, D.C., see Alugupalli, S. (3) 143
White, D.C., see Macnaughton, S.J. (1/2) 19
Wiegel, J., see Brill, J.A. (1/2) 29
Wimpee, M.H., see Macnaughton, S.J. (1/2) 19

Zbinden, R., Müller, F., Brun, F. and von Graevenitz, A., Detection of clumping factor-positive *Staphylococcus lugdunensis* by Staphaurex Plus® (1/2) 95

ELSEVIER

Journal of Microbiological Methods 31 (1998) 211–212

Journal of Microbiological Methods

Subject Index of Volume 31

Accelerated solvent extraction (1/2) 19
Acridine orange (3) 167
Active Bacteria (3) 99
Adhesins (1/2) 37
Aerobic conditions (3) 205
Assay for sporulation genes (1/2) 29

Bacillus/Clostridium subphylum (1/2) 29
Bacterial communities (3) 135
Bacteriophages (3) 205
Bacteroides fragilis (3) 205
*Bac*light (3) 167
Bartonella (1/2) 51
bfp (3) 201
Binary vector (1/2) 89
Blackleg vaccine (1/2) 83

Carboxy fluorescein diacetate-succinimidyl ester (3) 113
Cellulose decomposition (1/2) 9
Chlorate resistant (3) 175
Clostridium chauvoei (1/2) 83
Colony blot (3) 201
Corynebacterium diphtheriae (3) 127
Cotton strip assay (CSA) (1/2) 9
CTC (1/2) 1
^{14}C-MPN (3) 99
^{14}C-PLFA (3) 99

Derivative (3) 143
Detection by PCR-products, Southern hybridization (1/2) 29
Diffusion (1/2) 67
Digoxigenin probe (3) 201
DNA polymerase 1 (1/2) 89
dpaA/B (1/2) 29
DVC (1/2) 1

E. coli (1/2) 37
*eae*A (3) 201
EAF (3) 201
EC values (3) 159
ELISA (1/2) 83
Endospore-formation (1/2) 29
Enterobacter cloacae (1/2) 67
Enteropathogenic *Escherichia coli* (EPEC) (3) 201
Enumeration (1/2) 59

Flagellar antigen (1/2) 83
Flow cytometry (1/2) 75
Fungal volatiles (1/2) 45
Fusarium (1/2) 45
Fusarium moniliforme (3) 175
Fusarium proliferatum (3) 175

Gram staining (3) 185

Headspace (1/2) 45
Heavy metal toxicity (3) 159
Hollow fibre (1/2) 67
Humics (3) 193
Hyperosmotic stress (1/2) 1

Identification (1/2) 51
Immunomagnetic separation (1/2) 75
Injured cells (3) 167
Intracellular pH (3) 113

Kinetic growth inhibition assay (3) 159

Lipid biomarkers (1/2) 19
Listeria monocytogenes high gradient column (1/2) 75
LIVE/DEAD® *Bac*Light™ VIABILITY kit (1/2) 1
M. tuberculosis (3) 143

Mass spectrometry (3) 143
Membrane (1/2) 67
Membrane H^+-ATPase (3) 113
Methane (3) 99
Microcolony formation (3) 185
MPN (1/2) 59
Mycocerosic acids (3) 143
Neisseria (3) 151

Nitrate (3) 175
Nitrite (3) 175

Oxidative stress (1/2) 1
Oxrase® (3) 205

PCR (1/2) 37, 51, 59, (3) 135, 193
PCR–SSCP (3) 127
Pentachlorophenol (1/2) 59

Phenanthrene (3) 99
Phenotypic Characterization (3) 99
Phospholipid ester-linked fatty acids (PLFA) (3) 99
Plant root (1/2) 67
Plant transformation (1/2) 89
Plasmid cointegrates (1/2) 89
Principal components analysis (3) 135
Processed meats (3) 167
Pseudomonas putida (3) 159

Radioactive Fingerprinting (3) 99
Rapid method (1/2) 51
Restriction analysis (3) 135
Rhizosphere (3) 185, (1/2) 67
Ribotyping (3) 127
RRNA 16S–23S spacer region (3) 127
16S rRNA (3) 135

Saccharomyces cerevisiae (3) 113
Sephadex (3) 193
Silicone immobilization (3) 185
Soil (1/2) 59
Soil bacteria (3) 185
Solubilization (3) 151
Sorbic acid (3) 113
Sphingomonas sp (1/2) 59
spo0A (1/2) 29
Spun column (3) 193
ssp (1/2) 29
Staphaurex Plus® (1/2) 95
Staphylococcus aureus (1/2) 95
Staphylococcus lugdunensis (1/2) 95
Sulphate (3) 175

Taxonomy (1/2) 29
Tbps (3) 151
Transferrin-receptor (3) 151
Transmission electron microscopy (1/2) 75
TRFP (3) 135
Trichodiene (1/2) 45
Trichothecenes (1/2) 45

Uropathogenic (1/2) 37
UTI (1/2) 37

Vector exchange (1/2) 89
Viability staining (3) 167
Vibrio fischeri (3) 159
Virulence (1/2) 37
VNC *Escherichia coli* (1/2) 1

Weibull distribution (1/2) 9